

An Estuarine Benthic Index of Biotic Integrity (B-IBI) for Chesapeake Bay

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ABSTRACT: A multimetric benthic index of biotic integrity (B-IBI) was developed using data from five Chesapeake Bay sampling programs conducted between 1972 and 1991. Attributes of the index were selected by comparing the response of 17 candidate measures of benthic condition (metrics) between a set of minimally affected reference sites and at all other sites for which data were available. This procedure was conducted independently for each of seven habitats defined by salinity and substrate. Fifteen of the 17 candidate metrics differed significantly between reference sites and other sites for at least one habitat. No metric differed significantly in all seven habitats; however, four metrics, species diversity, abundance, biomass, and percent of abundance as pollution-indicative taxa, differed in six habitats. The index was calculated by scoring each selected metric as 5, 3, or 1 depending on whether its value at a site approximated, deviated slightly from, or deviated greatly from conditions at the best reference sites. Validation based on independent data collected between 1992 and 1994 indicated that the index correctly distinguished stressed sites from reference sites 93% of the time, with the highest validation rates occurring in high salinity habitats.

Introduction

Benthic invertebrates are used extensively as indicators of estuarine environmental status and trends because numerous studies have demonstrated that benthos respond predictably to many kinds of natural and anthropogenic stress (Pearson and Rosenberg 1978; Dauer 1993; Tapp et al. 1993; Wilson and Jeffrey 1994). Many characteristics of benthic assemblages make them useful indicators (Bilyard 1987), the most important of which are related to their exposure to stress and the diversity of their response. Exposure to hypoxia is typically greatest in near-bottom waters and anthropogenic contaminants often accumulate in sediments where benthos live. Benthic organisms generally have limited mobility and cannot avoid these adverse conditions (Wass 1967). This immobility is advantageous in environmental assessments be-

cause, unlike most pelagic fauna, benthic assemblages reflect local environmental conditions (Gray 1979). The structure of benthic assemblages responds to many kinds of stress because these assemblages typically include organisms with a wide range of physiological tolerances, feeding modes, and trophic interactions (Pearson and Rosenberg 1978; Rhoads et al. 1978; Boesch and Rosenberg 1981).

Although descriptions of benthic assemblages have been useful for characterizing environmental conditions and gradients at local scales, benthic assemblages have not yet been used to their full potential as indicators in regional-scale studies or studies of entire estuaries because the structure of benthic assemblages also reflects natural variation related to salinity, sediment type, latitude, and depth (Boesch 1973, 1977; Dauer et al. 1984, 1987; Holland et al. 1987; Schaffner et al. 1987; Snelgrove and Butman 1994; Heip and Craeymeersch 1995). Benthic assemblages rarely are used to assess ecological condition across habitats because of the difficulty in separating variation in the condition of the assemblage caused by habitat differ-

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TABLE 1. Data sources used in developing the Chesapeake Bay B-IBI.

Data Source	Time Period	Sampling Gear and Sampling Area (m ²)	Number of Summer Samples	References
Chesapeake Bay Benthic Monitoring Program, Maryland	1984–1994	Petite Ponar (0.025) Van Veen (0.1) Box Corer (0.022)	1,814	Ranasinghe et al. (1994a)
Chesapeake Bay Benthic Monitoring Program, Virginia	1985–1994	Box Corer (0.0184)	180	Dauer and Alden (1995)
EPA's Environmental Monitoring and Assessment Program (EMAP)	1990–1993	Van Veen (0.044)	293	Paul et al. (1992)
James River Study	1971–1972	Ponar (0.05)	28	Diaz (1989)
Wolf Trap	1987–1991	Box Corer (0.063)	4	Schaffner (unpublished data)

ences from variation caused by natural and anthropogenic stresses. In site-specific assessments of benthic assemblages, the problem of accounting for habitat-induced variation is minimized by using nearby reference sites from the same kind of habitat. Benthic assessments also are used frequently in studies of trends, where the problem of habitat-induced variation can be minimized by returning to the same site or area.

One approach used to integrate biotic responses and account for natural habitat variations in the freshwater environment is to define habitat-specific reference conditions (Karr 1991; Kerans and Karr 1994; DeShon 1995). This approach defines expected conditions at sites free of anthropogenic stress, and then assigns categorical values for various descriptive metrics by comparison with observations at reference sites. The result is a multi-metric index of biotic condition, frequently referred to as an index of biotic integrity (IBI). We borrowed from that approach to develop a benthic index of biotic integrity (B-IBI) for use with summer estuarine benthic communities of Chesapeake Bay.

Methods

Macrobenthic infaunal data from five Chesapeake Bay sampling programs, each of which used a 0.5-mm sieve and identified organisms to the lowest possible taxonomic level, were used to develop the B-IBI (Table 1). Taxonomic inconsistencies among programs were eliminated by cross-correlating the species lists, identifying differences in nomenclature, and consulting the taxonomist for each program to resolve discrepancies. Data collected before 1992 were used to calibrate the index; data collected between 1992 and 1994 were used to validate the index. If a site was sampled more than once during a summer, which we defined as July 15 through September 30, only the first sample was used in the calibration dataset.

Our analytical approach was similar to the one Karr et al. (1986) used to develop comparable indices for freshwater fish and involved four activi-

ties: selecting measures of benthic macroinvertebrate assemblages (referred to by Karr as metrics) that differ between references and degraded areas; identifying quantitative thresholds that differentiate between degraded and reference areas for each selected metric; combining metric scores to develop an index that identifies the condition of the assemblage at a site; and validating the index with an independent dataset.

Attributes of the index were selected by comparing response of the benthic assemblage between minimally affected reference sites and all other sites in our database. Reference sites were selected from our dataset by first eliminating sites within highly developed watersheds and locations near known point-source discharges. The remaining sites were examined to identify those where no contaminant exceeded Long et al.'s (1995) effects range-median (ER-M) concentration, total organic content of the sediment was less than 2%, and dissolved oxygen concentration was consistently high. Dissolved oxygen concentration was considered to be consistently high if the site had no recorded measurement below 2 ppm, 90% of the observations were above 3 ppm, and at least 80% of the measurements were above 5 ppm. Sites for which less than five independent oxygen measurements were available were selected as reference sites only if they were in a region classified by the Chesapeake Bay Program as being free of dissolved oxygen stress (Magnien et al. 1995). One of the sampling programs (EMAP) included a 10-d, acute sediment bioassay using the amphipod *Ampelisca abdita* for each site. Bioassay survival also had to exceed 80% of control survival for sites sampled by this program to be selected as reference sites.

Two criteria were used to compare metric values between reference sites and other sites in the calibration dataset. First, a Mann-Whitney U-test was used to test for differences in mean. Second, distributions were compared using a Kolmogorov-Smirnov test. The latter criterion was particularly important for the abundance and biomass metrics

TABLE 2. Mean values by habitat for each candidate metric at reference sites (top number) and at all other sites (bottom number). Shading in the Table: Top number boldface indicates pair is different by Mann-Whitney test; bottom number boldface indicates different by Kolmogorov-Smirnov test. N/A = data unavailable for that habitat.

	Tidal Freshwater	Oligohaline	Low Mesohaline	High Mesohaline Sand	High Mesohaline Mud	Polyhaline Sand	Polyhaline Mud
Species Diversity							
Shannon-Weiner	1.89 1.90	2.49 1.84	2.41 1.84	3.35 2.44	2.84 1.66	4.06 2.88	3.55 1.10
Productivity							
Abundance (# m ⁻²)	3,849 6,713	2,052 3,163	1,954 2,669	2,765 3,541	1,319 1,681	4,122 2,878	2,770 2,140
Biomass (g dry) (wt m ⁻²)	10.7 21.9	26.5 28.9	9.6 18.7	19.9 5.8	8.4 2.9	29.9 10.0	8.1 0.9
Species composition							
Percent of biomass as pollution-indicative taxa	24.5 N/A	0.8 1	0.9 8.4	2.8 N/A	9 55.4	2.3 6.3	5.9 24.8
Percent of abundance pollution-indicative taxa	29.4 45.9	22.4 27.8	2.4 19.1	11.6 19.3	26.5 48.7	12.1 32.4	22.9 58.9
Percent of biomass as pollution-sensitive taxa	18 N/A	89.1 66.9	83 26.9	78.9 N/A	65.3 20.1	71.2 66.1	63.2 40.4
Percent of abundance as pollution-sensitive taxa	12.8 4.7	44.8 28.3	30.6 25.7	40.3 20.8	29.5 10.3	48.5 29.3	39.9 8.4
Trophic composition							
Percent of abundance as carnivores or omnivores	15.1 10.1	11.8 15	22.9 18.3	37.4 23.8	23.1 15.6	35.3 32.8	41.8 19.9
Percent of abundance as deep deposit feeders	66.6 69.6	37.8 35.1	23.8 21.8	24.3 21.3	21.4 18	25.1 12.4	21.4 9.4
Percent of abundance as suspension feeders	6.5 11.3	16.7 13.6	1.2 8.3	7.0 15.4	4.8 6.4	10.6 9.0	3.1 4.9
Percent of abundance as interface feeders	10.4 8.4	33.7 35.2	52.1 48.1	31.4 39.4	50.8 46.7	29.0 35.3	33.7 39.1
Depth distribution below sediment-water interface							
Percent of taxa deeper than 5 cm	31.4 N/A	20.0 4.2	37.6 18.1	26.8 N/A	35.1 19.9	47.6 47.1	35.5 15.2
Percent of taxa deeper than 10 cm	8.7 N/A	6.7 0	20.0 6.9	8.2 N/A	10.7 3.3	22.0 30.1	9.3 6.1
Percent of abundance deeper than 5 cm	11.1 N/A	8.6 0.7	27.7 8.9	11.7 N/A	21.4 5.2	34.9 32	16.6 7
Percent of abundance deeper than 10 cm	1.9 N/A	1.5 0	8.0 1.1	2.4 N/A	5.6 0.8	16.6 16.3	4.2 2.4
Percent of biomass deeper than 5 cm	14.1 N/A	10.0 3	68.8 17	63.6 N/A	61.6 11.5	58.3 68.5	42 17.4
Percent of biomass deeper than 10 cm	3.3 N/A	1.7 0	48.3 3	31.6 N/A	29.9 1	28.5 33.8	24.9 10.6

because the anticipated response at degraded sites could be higher or lower than at reference sites, depending on the severity of the stress.

We tested 17 candidate metrics representing measures of species diversity, productivity, species composition, depth distribution, and trophic composition (Table 2). To ensure that the index would be applicable to a wide array of datasets, we attempted to identify metrics that have been shown to be least sensitive to collection gear (Ewing et al. 1988). Metrics based upon individual species were avoided in favor of assemblage measures, which can be equally sensitive to the effects of pollution

but less sensitive to small changes in habitat, such as salinity and substrate type (Warwick 1988).

Species were classified into feeding guilds using literature descriptions of feeding behavior (Jorgensen 1966; Bousfield 1973; Fauchald and Jumars 1979; Dauer et al. 1981); a complete list of taxa included in each guild is available in Ranasinghe et al. (1994b). Lists of pollution-indicative (Table 3) and pollution-sensitive (Table 4) taxa were developed using a two-step procedure. First, candidate taxa were identified based on known opportunistic or equilibrium life-history characteristics (Boesch 1973; Grassle and Grassle 1974; McCall

TABLE 3. Taxa defined as pollution-indicative in the Chesapeake Bay B-IBI.

Annelida: Polychaeta	Annelida: Oligochaeta
<i>Asabellides oculata</i>	<i>Aulodrilus limnobius</i>
<i>Capitella</i> spp.	<i>Aulodrilus paucichaeta</i>
<i>Hypereteone heteropoda</i>	<i>Aulodrilus pigueti</i>
<i>Leitoscoloplos fragilis</i>	<i>Aulodrilus plurisetia</i>
<i>Paraprionospio pinnata</i>	<i>Bothrioneurum vejdoskyanum</i>
<i>Streblospio benedicti</i>	<i>Haber</i> cf. <i>speciosus</i>
Arthropoda: Insecta	<i>Isochaetides curvosetosus</i>
<i>Chironomus</i> spp.	<i>Isochaetides freyi</i>
<i>Cladotanytarsus</i> spp.	<i>Limnodrilus hoffmeisteri</i>
<i>Coelotanytus</i> spp.	<i>Potamothenix vejdoskyi</i>
<i>Glyptotendipes</i> spp.	<i>Quistadrius multisetosus</i>
<i>Polydora tripodura</i>	Tubificid immatures without capilliform chaetae
<i>Procladius subletti</i>	Mollusca: Bivalvia
<i>Tanytus</i> spp.	<i>Mulinia lateralis</i>
	<i>Nucula proxima</i>

1977; Rhoads et al. 1978; Gray 1979; Rhoads and Boyer 1982; Warwick 1986; Dauer 1993). Second, the list was verified and amended by comparing the abundance of each taxa at the reference sites with abundance at sites from the calibration dataset that had known pollution stress. Sites in the calibration dataset were considered to be pollution-stressed if any sediment contaminant exceeded the Long et al. (1995) ER-M concentration and survival in sediment toxicity tests was less than 80% of control (where toxicity data were available), or total organic carbon in the sediment exceeded 3%, or dissolved oxygen content was low. A site was considered to have low dissolved oxygen if the measured concentration was below 0.5 ppm on any occasion, or below 2 ppm on at least three different dates within a year.

Metrics were selected separately for each of seven habitats (Table 5). The seven habitat strata were established by cluster analysis (Bray-Curtis similarity coefficient; flexible sorting; $\beta = -0.25$) of the calibration dataset to identify dominant assemblages in Chesapeake Bay, and by analysis of variance to determine whether salinity, grain size, depth, and latitude differed significantly among groups of sites defined by the different assemblages (Ranasinghe et al. in preparation).

Thresholds for the selected metrics were based on the distribution of values for the metric at the reference sites. The IBI approach involves scoring each metric as 5, 3, or 1, depending on whether its value at a site approximates, deviates slightly, or deviates greatly from conditions at reference sites (Karr et al. 1986). Threshold values were established as approximately the 5th and 50th (median) percentile values for reference sites in each habitat. For each metric, values below the 5th percentile were scored as 1; values between the 5th and

TABLE 4. Taxa defined as pollution-sensitive in the Chesapeake Bay B-IBI.

Coelenterata: Anthozoa	Mollusca: Bivalvia
<i>Ceriantheopsis americanus</i>	<i>Anadara ovalis</i>
Annelida: Polychaeta	<i>Anadara transversa</i>
<i>Asychis elongata</i>	<i>Cyrtopleura costata</i>
<i>Bhawania heteroseta</i>	<i>Dosinia discus</i>
<i>Chaetopterus variopedatus</i>	<i>Ensis directus</i>
<i>Clumenella toruata</i>	<i>Macoma balthica</i>
<i>Diopatra cuprea</i>	<i>Mercenaria mercenaria</i>
<i>Glycera americana</i>	<i>Mya arenaria</i>
<i>Glycinde solitaria</i>	<i>Rangia cuneata</i>
<i>Loimia medusa</i>	<i>Spisula solidissima</i>
<i>Macroclimene zonalis</i>	<i>Tagelus divinus</i>
<i>Marenzelleria viridis</i>	<i>Tagelus plebeius</i>
<i>Mediomastus ambiseta</i>	<i>Tellina agilis</i>
<i>Nephtys picta</i>	Echinodermata: Ophiuroidea
<i>Spiochaetopterus costarum</i>	<i>Microphiopholis atra</i>
<i>Spiophanes bombyx</i>	
Arthropoda	
<i>Alpheus heterochaelis</i>	
<i>Biffarius bifurmis</i>	
<i>Callinassa setimanus</i>	
<i>Cyathura polita</i>	
<i>Listriella clumenellae</i>	
<i>Squilla empusa</i>	

50th percentiles were scored as 3, and values above the 50th percentile were scored as 5. Metric scores were combined into an index by computing the mean score across all metrics for which thresholds were developed. Assemblages with an average score less than three are considered stressed, as they have metric values that on average are less than values at the poorest reference sites.

Two of the metrics, abundance and biomass, respond bimodally; that is, the response can be greater than at reference sites with moderate degrees of stress and less than at reference sites with higher degrees of stress (Pearson and Rosenberg 1978; Dauer and Conner 1980; Ferraro et al. 1991). For these metrics, the scoring was modified so that both exceptionally high (those exceeding the 95th percentile at reference sites) and low (<5th percentile) responses were scored as a 1. Values between the 5th and 25th percentiles or between the

TABLE 5. Habitat definitions used in the Chesapeake Bay B-IBI (N/A = not applicable).

Habitat	Bottom Salinity (‰)	Silt-clay (<63 μ) Content by Weight (%)
Tidal freshwater	0–0.5	N/A
Oligohaline	≥ 0.5 –5	N/A
Low Mesohaline	≥ 5 –12	N/A
High Mesohaline sand	≥ 12 –18	0–40
High Mesohaline mud	≥ 12 –18	>40
Polyhaline sand	≥ 18	0–40
Polyhaline mud	≥ 18	>40

75th and 95th percentiles were scored as 3, and values between the 25th and 75th percentiles of the values at reference sites were scored as 5.

The index was validated by examining its response at a new set of reference sites and a new set of sites with known environmental stress. Data used for validation were collected between 1992 and 1994 and were independent of data used to calibrate the index. Criteria for defining reference sites and known stressed sites from the validation dataset were the same as those described for the calibration dataset.

The index was further validated by examining the consistency of the index response at sites that were sampled more than once during a summer. This test, suggested by Stewart and Loar (1994) as a prerequisite for biological indices, examines index stability assuming that the quality of the site should not change appreciably during the index period.

Results

One hundred fourteen sites from the Chesapeake Bay database met our criteria as reference sites for developing the B-IBI. At least 10 reference sites were identified from each habitat except the oligohaline habitat, for which only seven reference sites were available. Biomass and depth metrics could not be evaluated in the tidal freshwater and high mesohaline sand habitats, though, because not all sampling programs partitioned samples by depth within the sediment or measured biomass individually by taxonomic groups.

Fifteen of the 17 candidate metrics differed significantly in mean or in distribution between reference sites and all other sites in the calibration dataset for at least one habitat (Table 2). No metric differed significantly in all seven habitats; however, four metrics, species diversity, abundance, biomass and percent of abundance as pollution-indicative taxa, differed in six habitats. The number of metrics that differed significantly between reference sites and all other sites was considerably greater in high salinity than in low salinity (Table 2). Fourteen of the 17 metrics differed significantly at reference sites in polyhaline mud compared with only three in tidal freshwater, and one in the oligohaline habitat.

Table 6 lists the selected metrics and their thresholds. We incorporated each metric that differed significantly in six habitats into the index for all habitats because in each case the direction of the response was the same in the seventh habitat and in most cases the difference was significant at a marginally lower alpha level. We also limited the number of redundant metrics within a habitat to avoid over-weighting individual properties of the

assemblage. For instance, we selected either abundance-based or biomass-based metrics of species composition, but not both. We also limited the index to a single depth-distribution metric, and single trophic composition metric, within each habitat.

The B-IBI classified 93% of the validation sites correctly (Table 7). The lowest classification efficiency by habitat was 50% in high mesohaline sand, but this was based on only two samples. Classification efficiency was generally poorer in low salinities. Cumulative classification efficiency for habitats with salinity less than 18‰ was 84%, whereas classification efficiency was 97% for salinities above 18‰ (Table 7).

The index was stable within the summer index period, with an overall correlation of 0.97 between the first and second sampling visits to a site. The correlation between the first and second sampling visits was greater than 0.9 in all habitats, and for six habitats the correlation exceeded 0.95 (Table 8). In all habitats except the oligohaline and mesohaline sand, more than 80% of the sites sampled twice during the same summer classified the same (Table 8). Most of the sites that classified differently between the two visits were sites for which index values were similar for both visits but were close to, and on either side of, the threshold value of 3.

Discussion

Development of the B-IBI was based upon quantifying established principles of benthic ecology, not on developing new ones. The principles were based largely on the paradigm of Pearson and Rosenberg (1978), which holds that benthic assemblages respond to improvements in habitat quality in three progressive stages: the abundance of organisms increases; species diversity increases as new taxa are able to survive at the site; and the dominant species at the site change from pollution-tolerant to pollution-sensitive species. In our work, this paradigm was supplemented with assumptions that the abundance and diversity of species occurring deeper in the sediment should be greater at reference sites than at degraded sites (Warwick 1986; Schaffner 1990; Dauer 1993) and that the distribution of benthos among feeding guilds should be more diverse at reference sites (Word 1978).

Each of these principles was successful at distinguishing reference sites from degraded sites within at least some habitats of Chesapeake Bay (Table 2). When applied individually to the validation dataset, however, none of the metrics derived from these principles were as effective at distinguishing between reference sites and degraded sites as the

TABLE 6. Thresholds used to score each metric of the Chesapeake Bay B-IBI.

	Scoring Criteria		
	5	3	1
Tidal Freshwater			
Shannon-Weiner	≥ 1.8	1.0–1.8	< 1.0
Abundance (# m ⁻²)	$\geq 1,000$ –4,000	500–1,000 or $\geq 4,000$ –10,000	< 500 or $\geq 10,000$
Biomass (g m ⁻²)	≥ 0.5 –3	0.25–0.5 or ≥ 3 –50	< 0.25 or ≥ 50
Abundance of pollution-indicative taxa (%)	≤ 25	25–75	> 75
Oligohaline			
Shannon-Weiner	≥ 2.5	1.9–2.5	< 1.9
Abundance (# m ⁻²)	$\geq 1,500$ –3,000	500–1,500 or $\geq 3,000$ –8,000	< 500 or $\geq 8,000$
Biomass (g m ⁻²)	≥ 3 –25	0.5–3 or ≥ 25 –60	< 0.5 or ≥ 60
Abundance of pollution-indicative taxa (%)	≤ 25	25–75	> 75
Abundance of pollution-sensitive taxa (%)	≥ 40	10–40	< 10
Low Mesohaline			
Shannon-Weiner	≥ 2.5	1.7–2.5	< 1.7
Abundance (# m ⁻²)	$\geq 1,500$ –2,500	500–1,500 or $\geq 2,500$ –6,000	< 500 or $\geq 6,000$
Biomass (g m ⁻²)	≥ 5 –10	1–5 or ≥ 10 –30	< 1 or ≥ 30
Abundance of pollution-indicative taxa (%)	≤ 10	10–20	> 20
Biomass of pollution-sensitive taxa (%)	≥ 80	40–80	< 40
Biomass >5 cm below sediment-water interface (%)	≥ 80	10–80	< 10
High Mesohaline sand			
Shannon-Weiner	≥ 3.2	2.5–3.2	< 2.5
Abundance (# m ⁻²)	$\geq 1,500$ –3,000	1,000–1,500 or $\geq 3,000$ –5,000	$< 1,000$ or $\geq 5,000$
Biomass (g m ⁻²)	≥ 3 –15	1–3 or ≥ 15 –50	< 1 or ≥ 50
Abundance of pollution-indicative taxa (%)	≤ 10	10–25	> 25
Abundance of pollution-sensitive taxa (%)	≥ 40	10–40	< 10
Abundance of carnivores and omnivores (%)	≥ 35	20–35	< 20
High Mesohaline mud			
Shannon-Weiner	≥ 3.0	2.0–3.0	< 2.0
Abundance (# m ⁻²)	$\geq 1,500$ –2,500	1,000–1,500 or $\geq 2,500$ –5,000	$< 1,000$ or $\geq 5,000$
Biomass	≥ 2 –10	0.5–2 or ≥ 10 –50	$< 1,000$ or $\geq 5,000$
Biomass of pollution-indicative taxa (%)	≤ 5	5–30	> 30
Biomass of pollution-sensitive taxa (%)	≥ 60	30–60	< 30
Abundance of carnivores and omnivores (%)	≤ 25	10–25	< 10
Biomass >5 cm below sediment-water interface (%)	≥ 60	10–60	< 10
Polyhaline sand			
Shannon-Weiner	≥ 3 –5	2.7–3.5	< 2.7
Abundance (# m ⁻²)	$\geq 3,000$ –5,000	1,500–3,000 or $\geq 5,000$ –8,000	$< 1,500$ or $\geq 8,000$
Biomass (g m ⁻²)	≥ 5 –20	1–5 or ≥ 20 –50	< 1 or ≥ 50
Biomass of pollution-indicative taxa (%)	≤ 5	5–15	> 15
Abundance of pollution-sensitive taxa (%)	≥ 50	25–50	< 25
Abundance of deep-deposit feeders (%)	> 25	10–25	< 10
Polyhaline mud			
Shannon-Weiner	≥ 3.3	2.4–3.3	< 2.4
Abundance (# m ⁻²)	$\geq 1,500$ –3,000	1,000–1,500 or $\geq 3,000$ –8,000	$< 1,000$ or $\geq 8,000$
Biomass (g m ⁻²)	≥ 3 –10	0.5–3 or ≥ 10 –30	< 0.5 or ≥ 30
Biomass of pollution-indicative taxa (%)	≤ 5	5–20	> 20
Biomass of pollution-sensitive taxa (%)	≥ 60	30–60	< 30
Abundance of carnivores and omnivores (%)	≥ 40	25–40	< 25
Taxa >5 cm below sediment-water interface (%)	≥ 40	10–40	< 10

index that combines them (Table 9). We attribute this to the staged response of benthos to stress, in which different metrics display greater response with different degrees of perturbation (Pearson and Rosenberg 1978); this illustrates one of the strengths of a multimetric approach (Karr 1991).

The use of multimetric indices has been adopted quite widely for fish but only recently for benthos

(Kerans and Karr 1994). Developing indices for estuarine benthos is difficult because of the lack of information about the life history or pollution sensitivity of most species; such information is necessary to develop metrics such as percent of individuals as pollution-indicative or pollution-sensitive taxa. Controlled experimentation to assess the pollution tolerance of individual taxa, which are nu-

TABLE 7. Classification efficiency of the B-IBI for sites from the validation dataset.

Habitat Class	Number of Validation Samples	Percent Correctly Classified
Tidal freshwater	22	86.4
Oligohaline	5	80.0
Low Mesohaline	29	82.3
High Mesohaline sand	2	50.0
High Mesohaline mud	50	96.0
Polyhaline sand	11	100.0
Polyhaline mud	52	100.0

merous for fish, are very limited for benthos. The pollution tolerance of benthos typically is categorized based on their life-history characteristics (Dauer 1993), which in some cases are ambiguous or inconsistent with their response to pollution (Seitz and Schaffner 1995).

One example of this conflict is our classification of the capitellid polychaete *Mediomastus ambiseta* as a pollution-sensitive taxon. Most previous studies have identified this widespread species as opportunistic and characteristic of pollution-stressed sites (Grassle and Grassle 1984; Dauer 1993). Our comparison of its abundance at reference sites and anthropogenically stressed sites showed the opposite (Table 10). We chose to rely on the empirical data from Chesapeake Bay in developing our index, but suggest that the classification of taxa into such groups is a fruitful area for consistent, controlled experimentation.

Although the principles on which the B-IBI is based were effective for distinguishing between sites of high and low quality, they did not apply equally well in all habitats. The number of metrics that differed significantly between sites of high and low quality in the calibration dataset was considerably smaller in the tidal freshwater and oligohaline habitat, and index validation efficiency was progressively weaker at lower salinities. Furthermore, our validation for the tidal freshwater habitat was not rigorous because our validation dataset did not include any degraded sites, and the threshold values we identified for tidal freshwater were low relative to those used for the other habitats. We suggest caution in using the tidal freshwater component of the B-IBI until further validation with a more complete dataset can be accomplished.

The lower validation efficiency for the index as a whole and the smaller number of metrics that worked individually in low salinities may be due to the inherently different fauna that inhabit the low salinity habitats (i.e., dominated by oligochaetes and insects instead of polychaetes, bivalves, and crustaceans). The principles of benthic ecology used to generate the list of candidate metrics were

TABLE 8. Classification consistency for sites visited more than once during a year.

Habitat Class	Number of Site-Year Combinations with Multiple Summer Samples	Percent of Sites with Unchanged Status	Correlation Between RGI Values
Tidal freshwater	19	84.2	0.98
Oligohaline	34	76.5	0.97
Low Mesohaline	103	92.2	0.98
High Mesohaline sand	74	66.3	0.95
High Mesohaline mud	60	85.0	0.96
Polyhaline sand	2	100.0	1.00
Polyhaline mud	35	97.1	0.91

based primarily on studies conducted in high salinity habitats (e.g., Pearson and Rosenberg 1978) and may not be entirely applicable to tidal freshwater benthic communities. The principles developed for nontidal freshwater environments (Kerans and Karr 1994; Lenat and Barbour 1994) differ substantially from those used in estuaries.

One constraint upon using the B-IBI is that it includes several metrics that were not measured by all benthic sampling programs in Chesapeake Bay. Many historic Chesapeake Bay collection efforts did not measure biomass, and few ongoing efforts partition the samples by depth, as is required for metrics in the higher salinity strata. To minimize the effect of missing biomass data, abundance-based substitutes for the pollution-indicative and pollution-sensitive categories are given (Table 11) because these metrics were also significant in all habitats where biomass-based metrics were selected for inclusion in the index. To assess the effect of missing data on reliability of the index, we repeated the validation step without using biomass data, did the same without the depth-related data, and again without either of these. Our overall classification efficiency when biomass metrics were removed fell from 93% to 92%. Dropping the depth-

TABLE 9. Classification efficiency for each of the metrics used in the B-IBI based on the validation data set. Metric testing was limited to habitats for which thresholds were established.

	Percent of Sites Correctly Classified
Shannon-Weiner diversity	89.5
Abundance	82.5
Biomass	81.6
Percent of abundance as pollution-indicative taxa	72.1
Percent of biomass as pollution-indicative taxa	87.5
Percent of abundance as pollution-sensitive taxa	90.2
Percent of biomass as pollution-sensitive taxa	82.4
Percent of abundance as carnivores and omnivores	71.2
Percent of abundance as deep deposit feeders	90.9
Percent of biomass deeper than 5 cm	90.9
Percent of taxa deeper than 5 cm	65.7

TABLE 10. Comparison of abundance and frequency of occurrence of *Mediomastus ambiseta* at reference and degraded sites in all habitats in which they were collected.

Habitat	Dataset	Abundance (number m ⁻²)		Frequency of Occurrence (Percent of Sites)	
		Reference Sites	Degraded Sites	Reference Sites	Degraded Sites
Polyhaline sand	Calibration	1,004	0	72	0
Polyhaline mud	Calibration	693	15	58	4
High Mesohaline sand	Calibration	849	23	58	46
High Mesohaline mud	Calibration	409	9	30	3
Polyhaline sand	Validation	2,073	0	77	0
Polyhaline mud	Validation	2,287	77	71	10
High Mesohaline mud	Validation	2,221	26	66	9

related metrics also resulted in a reduction in classification efficiency from 93% to 92%. Removing biomass and depth-related metrics together caused no further loss in classification efficiency.

In examining the validation results, we discovered a small number of instances in which just a few individuals caused the pollution-sensitive taxa metric to be scored as 5 when overall abundance in the sample was low. In such cases, a few stray organisms can artificially inflate the response. To avoid this, we recommend assigning a maximum score of 3 for the pollution-sensitive taxa metric if overall abundance at the site is less than at the reference sites (i.e., the 5th percentile IBI abundance threshold) for the habitat.

Our validation test showed a high overall classification efficiency, but it did not address whether the index is equally sensitive to the many different types of anthropogenic stress occurring in Chesapeake Bay. To assess this, we repeated the validation step separately for several categories of stress, including hypoxia, chemical contamination, and

enrichment of total organic carbon (TOC), and combinations of these stresses. To increase the number of samples to address this question, we included samples from the calibration dataset meeting our criteria as anthropogenically stressed sites in the analysis since these sites were used to only a small degree in calibration. The B-IBI was most effective at identifying hypoxia; it correctly identified more than 95% of hypoxic sites regardless of whether hypoxia occurred alone or in combination with other stresses (Table 12). The index identified poor biotic conditions in the presence of contaminants about 90% of the time but slightly less often when combined with hypoxia or high TOC. Response to organic enrichment was poorest; the index identified only about 70% of the enriched sites as having degraded benthic communities. The index may have been less efficient at identifying organically enriched sites because changes in the benthic assemblage at low levels of TOC enrichment are too small to be detected consistently with community-level attributes (Gray et al. 1990). Alternatively, the decreased efficiency may be a function of choosing 3% TOC as a threshold for enrichment; the efficiency of the index increased to 100% as TOC levels exceeded 4%.

Some authors have suggested that hypoxic conditions in portions of Chesapeake Bay are caused by natural seasonal stratification of the water column rather than anthropogenic influences (Officer et al. 1984; Cooper and Brush 1993). Only a few of our sites were located in these deep, stratified areas of the bay, but we observed biotic response to hypoxia to be the same, regardless of the cause of the hypoxia. Thus, the index cannot be

TABLE 11. Abundance-based thresholds that may be substituted for biomass-based thresholds.

	Scoring Criteria		
	5	3	1
Low Mesohaline			
Abundance of pollution-sensitive taxa (%)	≥25	5–25	<5
High Mesohaline mud			
Abundance of pollution-indicative taxa (%)	≤5	5–30	>30
Abundance of pollution-sensitive taxa (%)	≥60	30–60	<30
Polyhaline sand			
Abundance of pollution-indicative taxa (%)	≤10	10–40	>40
Polyhaline mud			
Abundance of pollution-indicative taxa (%)	≤15	15–50	>50
Abundance of pollution-sensitive taxa (%)	≥40	25–40	<25

TABLE 12. Classification efficiency of B-IBI by stressor, alone and in combination with other stressors.

	Alone	In Combination
Toxic contaminants	92.9%	88.1%
Hypoxia	98.5%	96.4%
Organic enrichment	68.8%	77.3%

used to distinguish natural from anthropogenic stress.

Although factors like natural stress must be considered in interpreting results, the B-IBI allows scientists to analyze and interpret benthic data in ways not previously possible by providing a uniform scale for comparing the quality of the benthic assemblage across habitats. This kind of information can be used to identify areas of the bay most in need of management action or to evaluate progress toward a set of environmental goals. The B-IBI is an extension of an effort to establish benthic restoration goals for Chesapeake Bay (Ranasinghe et al. 1994b).

The B-IBI also facilitates communication of complex benthic data. Many tools used previously to summarize benthic data, such as graphical models (Warwick 1986; Dauer 1993) or multivariate techniques (Warwick and Clarke 1991; Norris 1995), are robust but require detailed explanations and complex analytical techniques (McManus and Pauly 1990). Some scientists have expressed concern about indices that summarize too much information into a single number (Elliot 1994). The simple additive scaling and equal weighting of metrics in the B-IBI allow users to examine its component parts easily and to identify how each metric contributes to the overall score. Also, use of the B-IBI does not preclude examining and interpreting species composition data, particularly for sites classified near the threshold of 3, if this is considered necessary, desirable, or expedient.

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LITERATURE CITED

- BILYARD, G. R. 1987. The value of benthic infauna in marine pollution monitoring studies. *Marine Pollution Bulletin* 18:581-585.
- BOESCH, D. F. 1973. Classification and community structure of macrobenthos in the Hampton Roads area, Virginia. *Marine Biology* 21:226-244.
- BOESCH, D. F. 1977. A new look at the zonation of benthos along the estuarine gradient, p. 245-266. In B. C. Coull (ed.), *Ecology of Marine Benthos*. University of South Carolina Press, Columbia, South Carolina.
- BOESCH, D. F. AND R. ROSENBERG. 1981. Response to stress in marine benthic communities, p. 179-200. In G. W. Barret and R. Rosenberg (eds.), *Stress Effects on Natural Ecosystems*. John Wiley & Sons, New York.
- BOUSFIELD, E. L. 1973. *Shallow-water Gammaridean Amphipoda of New England*. Cornell University Press, Ithaca, New York.
- COOPER, S. R. AND G. S. BRUSH. 1993. A 2500-year history of anoxia and eutrophication in Chesapeake Bay. *Estuaries* 16: 617-626.
- DAUER, D. M. 1993. Biological criteria, environmental health and estuarine macrobenthic community structure. *Marine Pollution Bulletin* 26:249-257.
- DAUER, D. M. AND R. W. ALDEN. 1995. Long-term trends in the macrobenthos and water quality of the lower Chesapeake Bay (1985-1991). *Marine Pollution Bulletin* 30:840-850.
- DAUER, D. M. AND W. G. CONNER. 1980. Effects of moderate sewage on benthic polychaete populations. *Estuarine and Coastal Marine Science* 10:335-346.
- DAUER, D. M., R. M. EWING, AND A. J. RODI, JR. 1987. Macrobenthic distribution within the sediment along an estuarine salinity gradient. *Internationale Revue der Gesamten Hydrobiologie* 72:529-538.
- DAUER, D. M., C. A. MAYBURY, AND R. M. EWING. 1981. Feeding behavior and general ecology of several spionid polychaetes from Chesapeake Bay. *Journal of Experimental Marine Biology and Ecology* 54:21-38.
- DAUER, D. M., T. L. STOKES, JR., H. R. BARKER, JR., R. M. EWING, AND J. W. SOURBEER. 1984. Macrobenthic communities of the lower Chesapeake Bay. IV. Baywide transects and the inner continental shelf. *Internationale Revue der Gesamten Hydrobiologie* 69:1-22.
- DESHON, J. E. 1995. Development and application of the invertebrate community index, p. 217-243. In W. P. Davis and T. P. Simon (eds.), *Biological Assessment and Criteria*. Lewis Publishers, Boca Raton, Florida.
- DIAZ, R. J. 1989. Pollution and tidal benthic communities of the James River Estuary, Virginia. *Hydrobiologia* 180:195-211.
- ELLIOTT, M. 1994. The analysis of macrobenthic community data. *Marine Pollution Bulletin* 28:62-64.
- EWING, R. M., J. A. RANASINGHE, AND D. M. DAUER. 1988. Comparison of five benthic sampling devices. Prepared for the Virginia Water Control Board, Richmond, Virginia.
- FAUCHALD, K. AND P. A. JUMARS. 1979. The diet of worms. A study of polychaete feeding guilds. *Oceanography and Marine Biology Annual Review* 17:193-284.
- FERRARO, S. P., R. C. SWARTZ, F. A. COLE, AND D. W. SCHULTS. 1991. Temporal changes in the benthos along a pollution gradient: Discriminating the effects of natural phenomena from sewage-industrial wastewater effects. *Estuarine Coastal and Shelf Science* 33:383-407.
- GRASSLE, J. F. AND J. P. GRASSLE. 1974. Opportunistic life histories and genetic systems in marine benthic polychaetes. *Journal of Marine Research* 32:253-284.
- GRASSLE, J. P. AND J. F. GRASSLE. 1984. The utility of studying the effects of pollutants on single species populations in benthos of mesocosms and coastal ecosystems, p. 621-642. In H. H. White (ed.), *Concepts in Marine Pollution Measurements*, Maryland Sea Grant College, College Park, Maryland.
- GRAY, J. S. 1979. Pollution-induced changes in populations. *Transactions of the Royal Philosophical Society of London (B)* 286: 545-561.
- GRAY, J. S., K. R. CLARKE, R. M. WARWICK, AND G. HOBBS. 1990. Detection of initial effects of pollution on marine benthos: An example from the Ekofisk and Eldfisk oilfields, North Sea. *Marine Ecology Progress Series* 66:285-299.
- HEIP, C. AND J. A. CRAEYMEERSCH. 1995. Benthic community structures in the North Sea. *Helgolander Meeresuntersuchungen* 49:313-328.
- HOLLAND, A. F., A. SHAUGHNESSY, AND M. H. HEIGEL. 1987.

- Long-term variation in mesohaline Chesapeake Bay benthos: Spatial and temporal patterns. *Estuaries* 10:227-245.
- JORGENSEN, C. B. 1966. Biology of Suspension Feeding. Pergamon Press, Oxford, England.
- KARR, J. R. 1991. Biological integrity: A long-neglected aspect of water resource management. *Ecological Applications* 1:66-84.
- KARR, J. R., K. D. FAUSCH, P. L. ANGERMEIER, P. R. YANT, AND I. J. SCHLOSSER. 1986. Assessing biological integrity in running waters: A method and its rationale. Special Publication 5. Illinois Natural History Survey, Champaign, Illinois.
- KERANS, B. L. AND J. R. KARR. 1994. A benthic index of biotic integrity (B-IBI) for rivers of the Tennessee Valley. *Ecological Applications* 4:768-785.
- LENAT, D. R. AND M. T. BARBOUR. 1994. Using benthic macroinvertebrate community structure for rapid, cost-effective, water-quality monitoring: Rapid bioassessment, p. 187-216. In S. L. Loeb and A. Spacie (eds.), *Biological Monitoring of Aquatic Systems*. Lewis Publishers, Boca Raton, Florida.
- LONG, E. R., D. D. McDONALD, S. L. SMITH, AND F. D. CALDER. 1995. Incidence of adverse biological effects within ranges of chemical concentrations in marine and estuarine sediments. *Environmental Management* 19:81-95.
- MAGNIEN, R., D. BOWARD, AND S. BIEBER. 1995. The state of the Chesapeake Bay 1995. Report by the United States Environmental Protection Agency Chesapeake Bay Program, Annapolis, Maryland.
- MCCALL, P. L. 1977. Community patterns and adaptive strategies of the infaunal benthos of Long Island Sound. *Journal of Marine Research* 35:221-266.
- MCMANUS, J. W. AND D. PAULY. 1990. Measuring ecological stress: Variations on a theme by R. M. Warwick. *Marine Biology* 106:305-308.
- NORRIS, R. H. 1995. Biological monitoring: The dilemma of data analysis. *Journal of the North American Biological Society* 14: 440-450.
- OFFICER, C. B., R. B. BIGGS, J. L. TAFT, L. E. CRONIN, M. A. TYLER, AND W. R. BOYNTON. 1984. Chesapeake Bay anoxia: Origin, development and significance. *Science* 223:22-27.
- PAUL, J. F., K. J. SCOTT, A. F. HOLLAND, S. B. WEISBERG, J. K. SUMMERS, AND A. ROBERTSON. 1992. The estuarine component of the U.S. EPA's Environmental Monitoring and Assessment Program. *Chemistry and Ecology* 7:93-116.
- PEARSON, T. H. AND R. ROSENBERG. 1978. Macrobenthic succession in relation to organic enrichment and pollution of the marine environment. *Oceanography and Marine Biology Annual Review* 16:229-311.
- RANASINGHE, J. A., L. C. SCOTT, R. NEWPORT, AND S. B. WEISBERG. 1994a. Chesapeake Bay Water Quality Monitoring Program, Long-Term Benthic Monitoring and Assessment Component. Level I Comprehensive Report, July 1984-December 1993. Prepared for the Maryland Department of the Environment, Baltimore, Maryland.
- RANASINGHE, J. A., S. B. WEISBERG, J. B. FRITHSEN, D. M. DAUER, L. C. SCHAFFNER, AND R. J. DIAZ. 1994b. Chesapeake Bay Benthic Community Restoration Goals. Report CBP/TRS 107/94. United States Environmental Protection Agency, Chesapeake Bay Program, Annapolis, Maryland.
- RHOADS, D. C. AND L. F. BOYER. 1982. The effects of marine benthos on physical properties of sediments: A successional perspective, p. 3-52. In P. L. McCall and M. S. Tevesz (eds.), *Animal-Sediment Relations*. Plenum Press, New York.
- RHOADS, D. C., P. L. MCCALL, AND J. Y. YINGST. 1978. Disturbance and production on the estuarine sea floor. *American Scientist* 66:577-586.
- SCHAFFNER, L. C. 1990. Small-scale organism distributions and patterns of species diversity: Evidence for positive interactions in an estuarine benthic community. *Marine Ecology Progress Series* 61:107-117.
- SCHAFFNER, L. C., R. J. DIAZ, C. R. OLSON, AND I. L. LARSEN. 1987. Faunal characteristics and sediment accumulation processes in the James River Estuary, Virginia. *Estuarine Coastal and Shelf Science* 25:211-226.
- SEITZ, R. D. AND L. C. SCHAFFNER. 1995. Population ecology and secondary production of the polychaete *Loimia medusa* (Terebellidae). *Marine Biology* 121:701-711.
- SNELGROVE, P. V. AND C. A. BUTMAN. 1994. Animal-sediment relationships revisited: Cause vs. effect. *Oceanography and Marine Biology Annual Review* 32:111-177.
- STEWART, A. J. AND J. M. LOAR. 1994. Spatial and temporal variation in biological monitoring data, p. 91-124. In S. L. Loeb and A. Spacie (eds.), *Biological Monitoring of Aquatic Systems*. Lewis Publishers, Boca Raton, Florida.
- TAPP, J. F., N. SHILLABEER, AND C. M. ASHMAN. 1993. Continued observation of the benthic fauna of the industrialised Tees estuary, 1979-1990. *Journal of Experimental Marine Biology and Ecology* 172:67-80.
- WARWICK, R. M. 1986. A new method for detecting pollution effects on marine macrobenthic communities. *Marine Biology* 92:557-562.
- WARWICK, R. M. 1988. The level of taxonomic discrimination required to detect pollution effects on marine macrobenthic communities. *Marine Pollution Bulletin* 19:259-268.
- WARWICK, R. M. AND K. R. CLARKE. 1991. A comparison of some methods for analysing changes in benthic community structure. *Journal of the Marine Biological Association of the United Kingdom* 71:225-244.
- WASS, M. L. 1967. Indicators of pollution, p. 271-183. In T. A. Olsen and F. Burgess (eds.), *Pollution and Marine Ecology*. John Wiley and Sons, New York.
- WILSON, J. G. AND D. W. JEFFREY. 1994. Benthic biological pollution indices in estuaries, p. 311-327. In J. M. Kramer (ed.), *Biomonitoring of Coastal Waters and Estuaries*. CRC Press, Boca Raton, Florida.
- WORD, J. Q. 1978. The infaunal trophic index, p. 19-39. In W. Bascom (ed.), *Coastal Water Research Project, Annual Report for the Year 1978*. Southern California Coastal Water Research Project, El Segundo, California.

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